FETAL CELLS & FF DNA IN MATERNAL BLOOD: the new era of prenatal diagnosis

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Prevalence of Trisomies 21, 18, 13

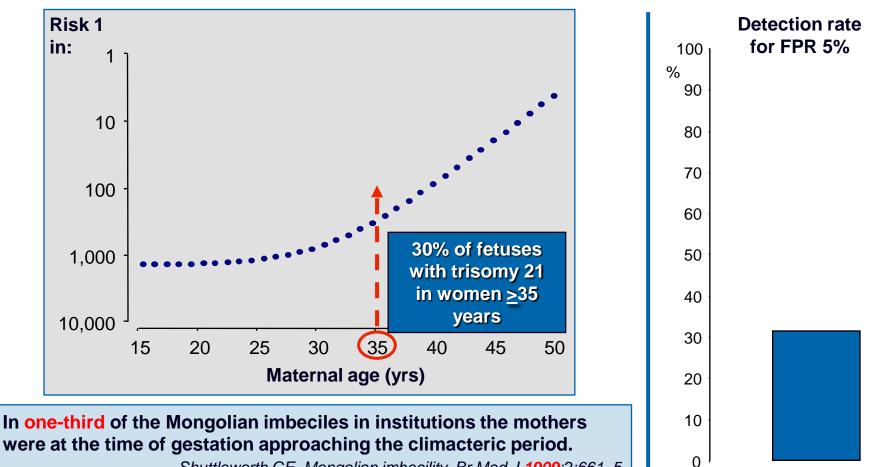
Trisomy Type	Condition Name	Frequency	
Chromosome 21	Down syndrome	1 in 700 live births	
Chromosome 18	Edwards syndrome	1 in 5,000 live births	
Chromosome 13	Patau syndrome	1 in 16,000 live births	

U.S. National Library of Medicine. Genetics Home Reference. Down Syndrome: http://ghr.nlm.nih.gov/condition/downsyndrome, Trisomy 18: http://ghr.nlm.nih.gov/condition/trisomy-18, Trisomy 13: http://ghr.nlm.nih.gov/condition/trisomy-13. Accessed July 12, 2012.

1970s

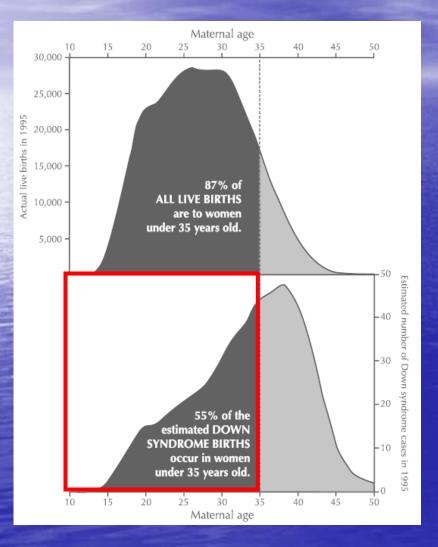
Screening for aneuploidies

Maternal age



Shuttleworth GE. Mongolian imbecility. Br Med J 1909;2:661–5

Importance of Screening All Pregnant Women



Majority of babies born with Down syndrome are in women under 35 years old

Provider handbook for The California Prenatal Screening Program 2009.

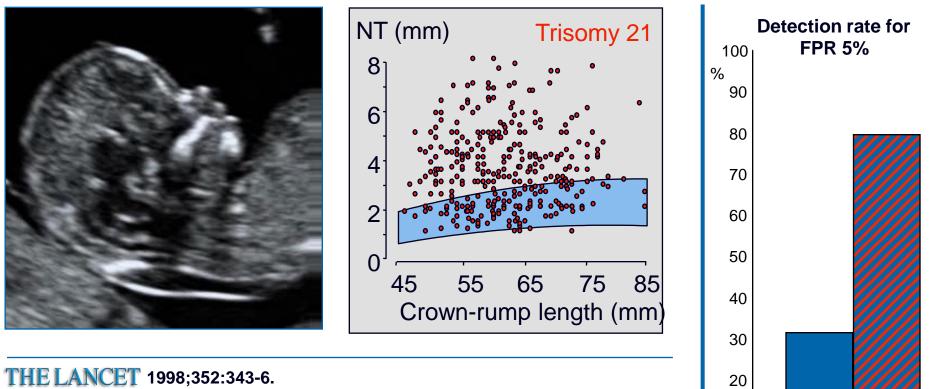
1990s

Screening for aneuploidies

10

0

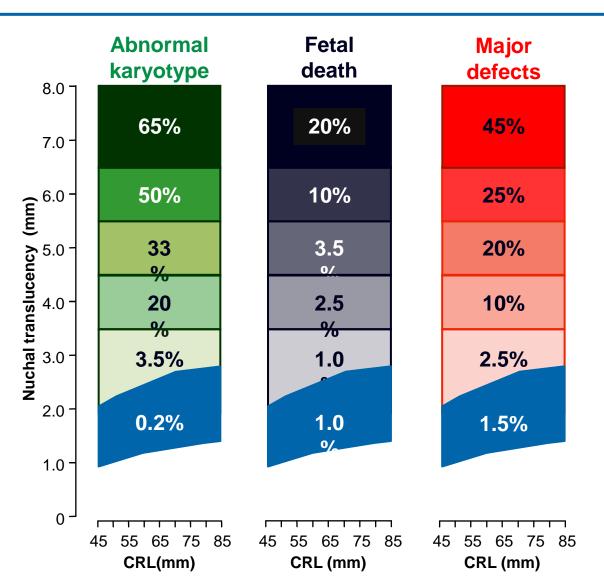
Maternal age and fetal nuchal translucency



Snijders RJ, Noble P, Sebire N, Souka A, Nicolaides KH Assessment of risk of trisomy 21 by maternal age and fetal nuchal-translucency thickness

96,127 singleton pregnancies, including 326 cases of trisomy 21: DR 77% for FPR 5%

Implications of increased NT



Cardiac defects Lethal skeletal dysplasias **Diaphragmatic hernia Exomphalos** Megacystis Akinesia deformation sequence Spinal muscular atrophy **Treacher-Collins syndrome** Jarcho-Levin syndrome **Beckwith-Wiedemman** syndrome Smith-Lemli-Opitz syndrome Zellweger syndrome Noonan syndrome di George syndrome **Congenital lymphedema** Dyserythropoietic anaemia Thalassaemia-a

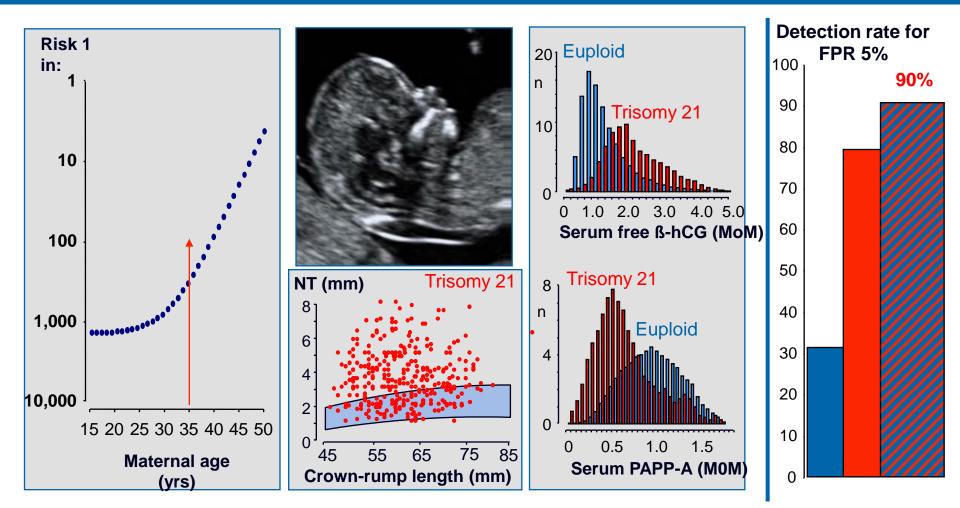
Parvovirus B19 infection

Souka et al. Am J Ob Gyn 2004

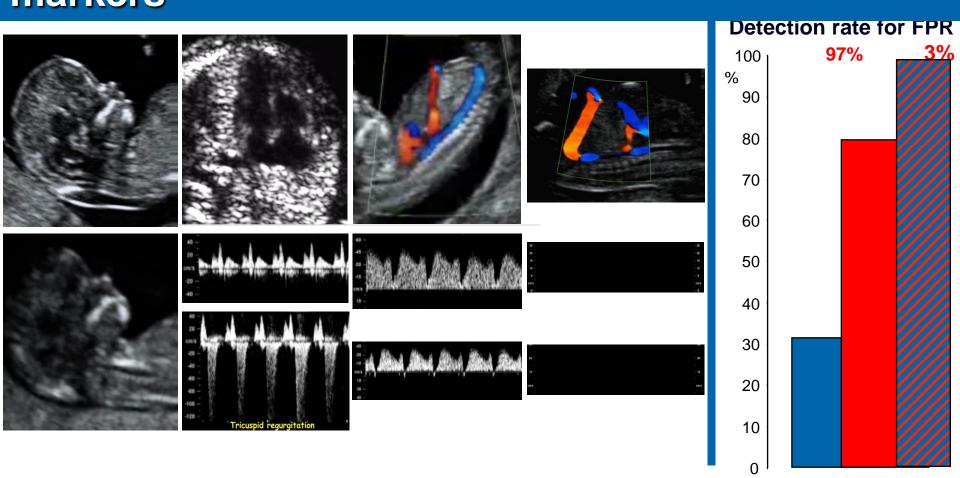
Screening for aneuploidies

1st trimester combined test

2000



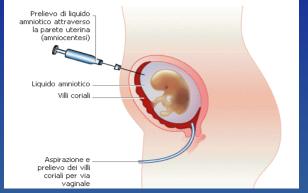
2000-10 Screening for aneuploidies 1st trimester combined test and additional US markers



CURRENT PRENATAL DIAGNOSIS TOOLS

INVASIVE Villocentesis Amniocentesis

Cordocentesis

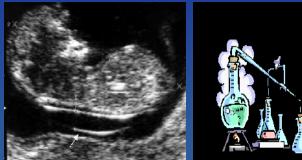


Offered to women at risk for:

- maternal age
- positive screening test
- chromosomal abnormalities
- previous affected child

NON INVASIVE

Ultrasound screening Biochemical screening



Low sensitivity and specificity (< 100%)

Potential Limitations of Current Screening Tests

High false positive rate (5%)

Late information Prolonged uncertainty Inconvenient Multiple visits Specialized ultrasound

Safety concerns

NEED TO DEVELOP NEW NON INVASIVE PRENATAL DIAGNOSTIC TESTS

SIMPLE
 EASY
 LEAST ANXIOUS
 MORE SENSITIVE
 LEAST AGGRESSIVE
 MORE SPECIFIC

NEW APPROACHES

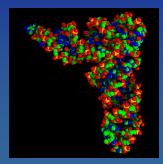
FETAL CELLS IN MATERNAL BLOOD

FREE FETAL DNA IN MATERNAL BLOOD





FREE FETAL RNA IN MATERNAL BLOOD





KEY BIOLOGICAL QUESTIONS

- Which is the ideal fetal cell type for a non invasive prenatal diagnosis?
- Which is the frequence of fetal cells in maternal blood?
- Which are suited laboratory approaches to enrich and to purify fetal cells in maternal blood?
- Are fetal cells always present in maternal blood during gestation?
- Are the fetal cells, isolated from maternal blood, sufficient for genetic diagnosis?
- Which is the best timing to retrieve fetal cells from maternal blood?



FETAL CELL TYPES IN MATERNAL BLOOD DURING GESTATION

Studies on fetal blood obtained by cordocentesis have been able to strengthen the knowledge of the composition and development of fetal blood component throughout pregnancy

✓ LYMPHOCYTES

✓ ERYTHROBLASTS

✓ TROPHOBLASTS

HEMATOPOIETIC STEM
 PROGENITOR CELLS

✓ MESENCHYMAL STEM CELLS

NUMBER OF FETAL CELLS

I fetal cell/ 10⁵ – 10⁸ maternal cells (Price JO et al., 1991; Hamada H et al., 1993; Langlois S et al., 1993)

> 1 fetal cell/ ml of maternal blood (Bianchi D et al., 1997)

> 2 -6 fetal cells/ml maternal blood (Krabchi et al., 2001)

0 – 2 fetal progenitor cells/ ml maternal blood (Guetta et al., 2003)

 Numerous studies demonstrated that in women carrying fetus with trisomy 21 or 13 and in pregnancies complicated by preeclampsia, the mean number of fetal cells increase in respect to normal pregnancies.
 (Holzgreve W et al., 2007)

TIME OF APPEARENCE OF FETAL CELLS IN MATERNAL CIRCULATION



After 40 days of gestation (Holzgreve W. et al., 1993)

✓ From 4th week of gestation (Peault B et al., 2003; Lo YMD et al., 1996)

11 - 16 weeks of gestation
 (Ideal time for isolating fetal cells from maternal blood)

HEMATOPOIETIC STEM PROGENITOR CELLS (HSPCs)



 Presence in maternal blood: HSPCs are present in maternal circulation from 4th weeks of gestation whereas their concentration decrease after 20 weeks.

- Identification: CD34, CD133 monoclonal antibodies;
- In vitro culture expansion has been studied and proposed by Lo et al. (Lancet, 1994), Little et al. (Blood 1997) and Di Renzo et al. (Journal of Hematotherapy & Stem Cell Research 2000).
- ✓ **Frequency:** fetal/maternal cell ratio is 1 per 4.75x10⁶- 1.6x10⁷ cells.
- ✓Advantages:
 - Clonogenicity;
 - Increased clonogenicity in fetal blood during early 2nd trimester;

- Versatility to culture and to proliferate extensively in vitro.



- ✓ Work in progress:
 - persistence in maternal blood after pregnancy: *solved*

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- new fetal HSPCs markers : *working on*

A NEW METHODOLOGY OF FETAL STEM CELL ISOLATION, PURIFICATION, AND EXPANSION: PRELIMINARY RESULTS FOR NON INVASIVE PRENATAL DIAGNOSIS

Tilesi, Coata, Di Renzo et al.

Journal of Hematotherapy & Stem Cell Research 2000; 9: 583-590



An enrichment of

33 times of BFU-E/CFU-E and

16 times of CFU-GM colonies after

miniMACS CD34+ HSPCs purification

was obtained

RESULTS

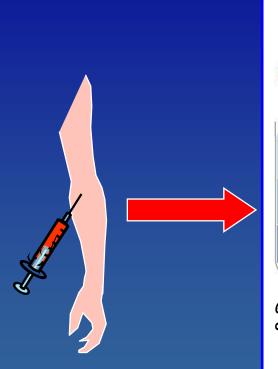
Results of FISH analysis with X and Y, 21 chromosome fluorescent probes in cultured cells

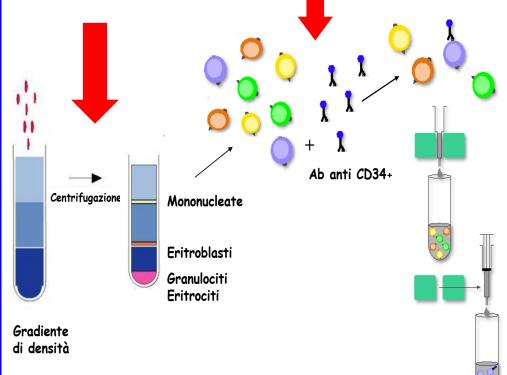
Slide Identific.	Fetal karyotype		Number cells with XY signals	cells	Number cells with trisomy 21 signals	Fetal/maternal cell ratio
*27	46, XY	669	5	-	_	1/133
19	46, XY	1433	6	-	-	1/238
† 3	46, XY	570	-	11	-	1/52
4	46, XY	1050	-	4	-	1/262
°18	47, XX+21	659	-	-	19	1/34
15	47, XY+21	100	-	-	8	1/12

IN SUMMARY.....

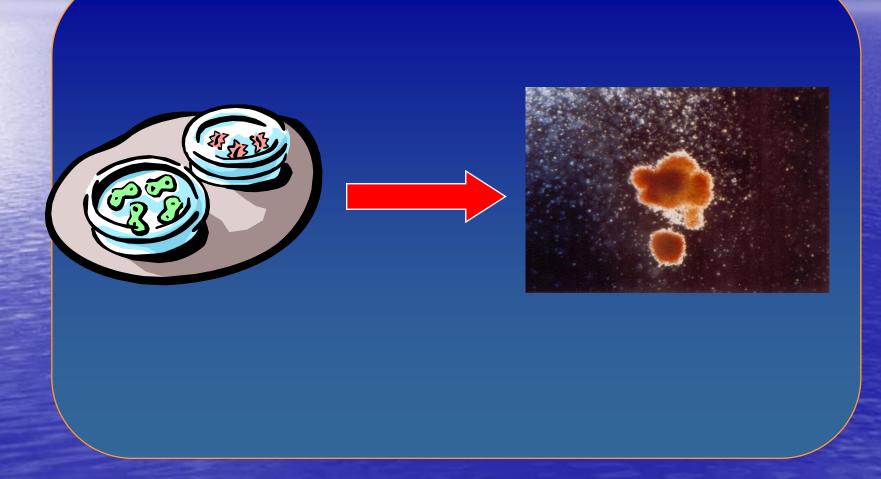
THE SAFE (SANGUE FETALE: FETAL BLOOD) TEST IS COMPRISING THREE STEPS

METHODOLOGY Selection of stem cells CD34+



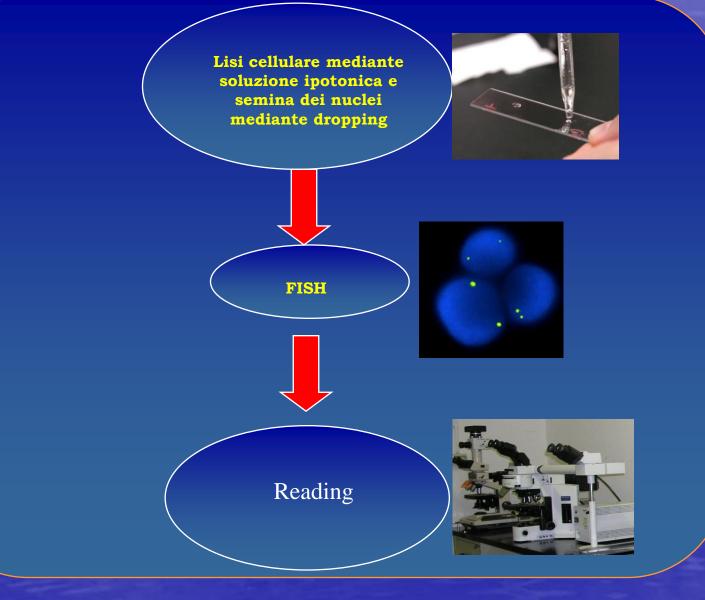


METHODOLOGY Expansion in vitro of CD 34+



METHODOLOGY

Preparation of nuclei by dropping and FISH



MOTORIZED MYCROSCOPE WITH AUTOMATED ACQUISITION SYSTEM

Microscope BX-61 Olympus with software BX-UCB Olympus



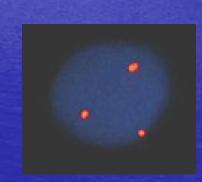
Objective changer

Motorized table with 4 sides of reading Fluorescence lamp (100Watt) at hight pression of mercury

FISH PERFORMED BY USING LSI 21 PROBE FOR THE NON INVASIVE DIAGNOSIS OF FETAL TRISOMY 21



A

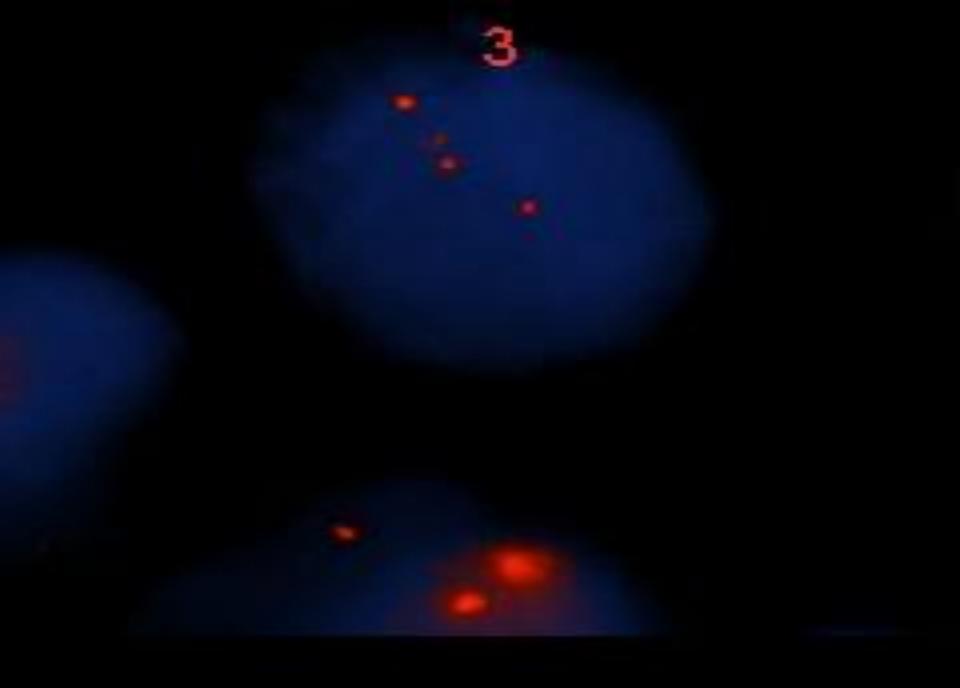


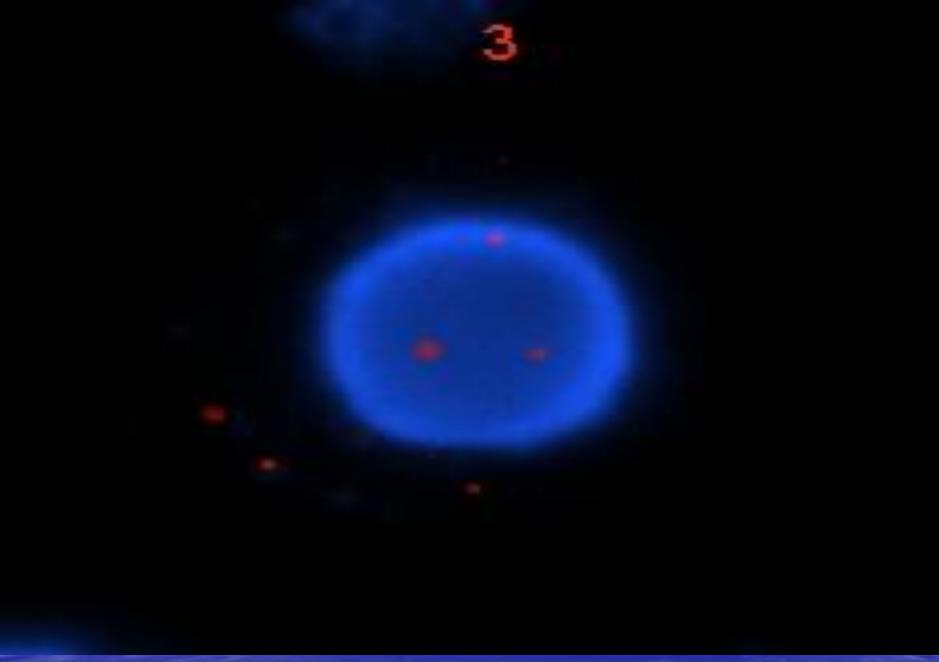
B

A and B: Frames obtained by using the automated mycroscope A: Two disomic nuclei for the chromosome 21 B: Fetal trisomic nucleus for the chromosome 21



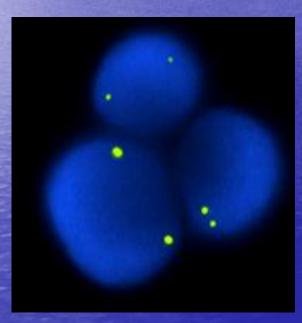
SAFE TEST: TRISOMY 21



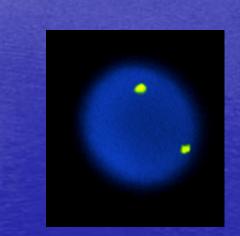


SAFE TEST: TRISOMY 21

FISH PERFORMED BY USING LSI 13 PROBE FOR THE NON INVASIVE DIAGNOSIS OF FETAL TRISOMY 13

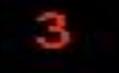


A



B

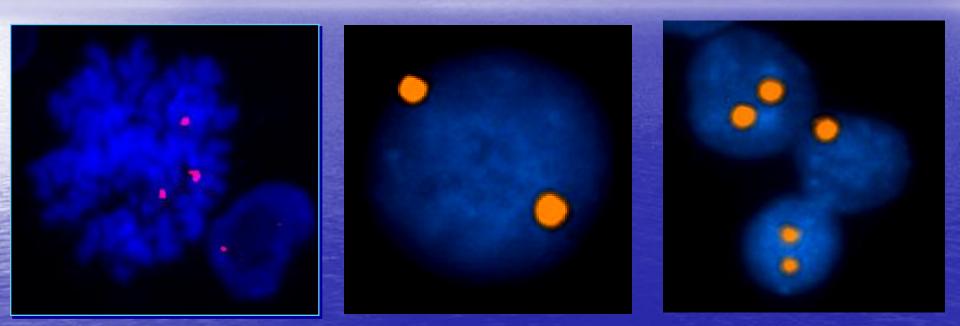
A and B: Frames obtained by using the automated mycroscope A and B: Each nucleus showed is disomic for the chromosome 13



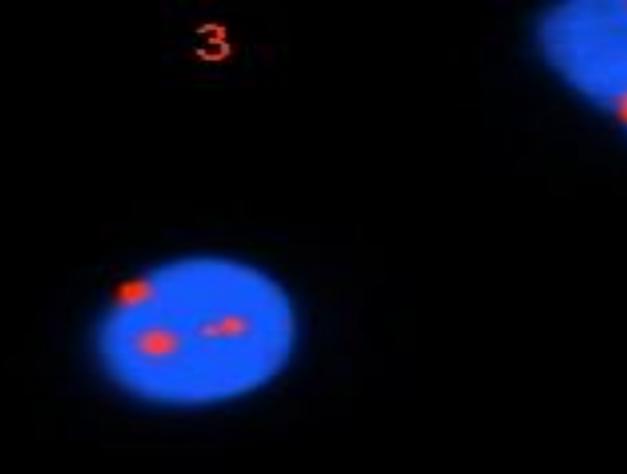
SAFE TEST: TRISOMY 13

FISH PERFORMED BY USING CEP 18 PROBE FOR THE NON INVASIVE DIAGNOSIS OF FETAL TRISOMY 18

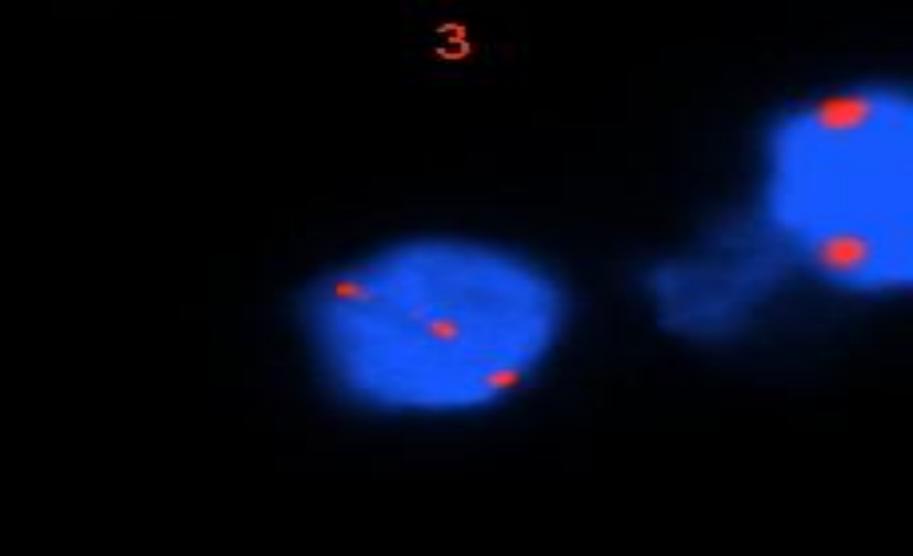
B



A: Fetal metaphase shows three 18 orange spots and a maternal nuclei with two 18 orange spots
B and C: Frames obtained by using the automated mycroscope
B: Disomic nucleus for the chromosome 18
C: Two disomic nuclei and one monosomic nucleus for the chromosome 18

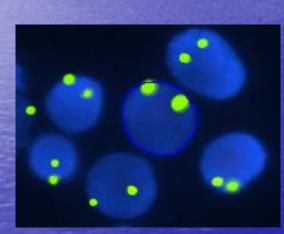


SAFE TEST: TRISOMY 18

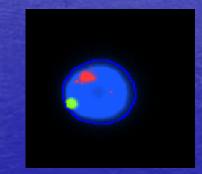


SAFE TEST: TRISOMY 18

FISH PERFORMED BY USING CEP XY PROBE FOR THE NON INVASIVE DIAGNOSIS OF FETAL GENDER



A

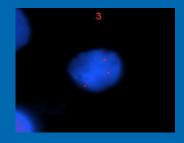


B

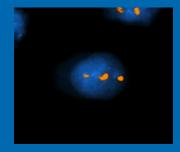
A and B: Frames obtained by using the automated mycroscope A: XX nuclei B: Fetal XY nucleus

SAFE TEST – FETAL CELLS

> 21 Chromosome analysis
Diagnostic accuracy 97.9%



18 Chromosome analysis Diagnostic accuracy 98.9%



> 13 Chromosome analysis Diagnostic accuracy 98.9%



RESULTS SAFE TEST 2006-2010

- 1782 tests: checked by CVS, amniocentesis, birth genetic map
- 18 trisomy 21
- 6 trisomy 18
- I trisomy 13
- 1 Klinefelter

Detection rate 100% Sensitivity 100%

Specificity 94%

• Chr 21 • Chr 18 • Chr 13 sens 100% • Chr X & Y sens 100%

sens 100% sens 100% **spec 91%** spec 92% spec ND spec 100%

NEW POSSIBILITIES FROM FREE FETAL DNA

Cell-free fetal DNA in maternal blood

THE LANCET

Presence of fetal DNA in maternal plasma and serum

1992

Dennis Lo et al. 1997;350:485

5% of total maternal plasma cfDNA is fetal

Fetal sex determination (X-linked disease)

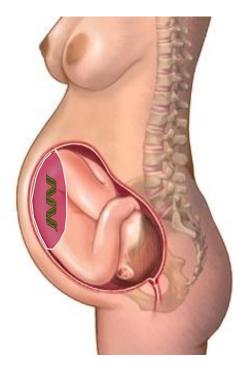
• Y chromosome in male fetuses

Hemolytic disease

• RHD gene in Rh D negative women

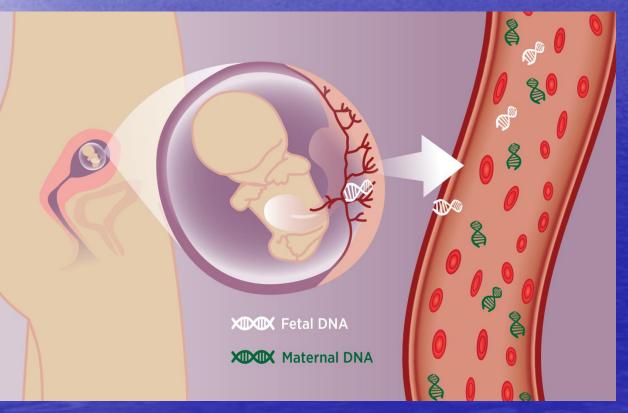
Autosomal dominant disease

 Achondroplasia, Myotonic dystrophy, Huntington's disease



Cell-free DNA in Maternal Blood

- Cell-free DNA (cfDNA) are short DNA fragments
- In pregnancy, cfDNA from both the mom and fetus are in maternal blood
- Amount of fetal cfDNA present is a small fraction of the maternal cfDNA



New possibilities: NIPT



Standard Blood Draw

- Simpler clinical protocol
- As early as 10 weeks gestation
- Higher detection rate
- 30-50x lower false positive rate

The Benefits

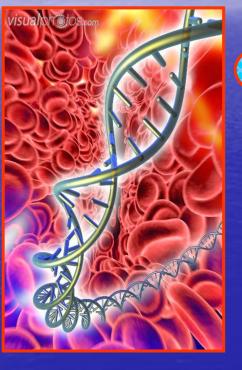
CELL-FREE FETAL DNA

Presence of cell-free fetal DNA in the maternal circulation

Fetal gender determination

Time: 10-13 weeks of gestation

Target population: pregnant women at risk of ambiguous genitalia, X-linked conditions and single gene disorders such as congenital adrenal hyperplasia



Fetal RhD genotyping

Time: from the 13th week of gestation

Target population: RhD-negative pregnant women

CELL-FREE FETAL DNA

DNA

BLOOD

SAMPLE



Density gradient centrifugation

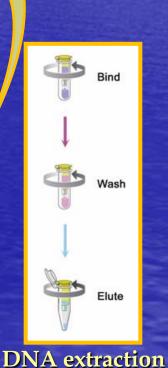


Peripheral blood sampling

METHODS

Analysis by real time PCR (Polymerase Chain Reaction)





PLASMAT

Non invasive fetal gender determination

Di Renzo et al. Prenat Diagn 2008 Am J Ob Gyn 2009 Clin Genet 2011

PRENATAL ASSESSMENT OF FETAL GENDER

- 1. So far, the test has been performed on 912 pregnant women .
- 2. The use of our interpretation criteria allowed us to improve the test by reducing false positive results.
- 3. The test is functional in clinical routine practice of non invasive prenatal diagnosis since it is easy, rapid and automated. After about 4 hours from the blood sampling it is possible to obtain the results of 20 samples simultaneously.

SENSITIVITY (%)	99.9
SPECIFICITY (%)	99.5
VPP (%)	99.5
VPN (%)	100
EFFICIENCY (%)	99.7

COMPARISON OF TWO DNA EXTRACTION METHODS FOR THE DEVELOPMENT OF A PRENATAL NONINVASIVE GENETIC TEST FOR FETAL STATUS RhD DIAGNOSIS

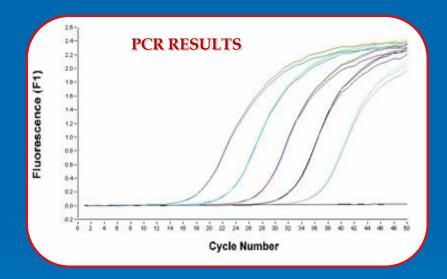
Fanetti, Coata, Di Renzo et al.

PRENAT DIAGN 2010



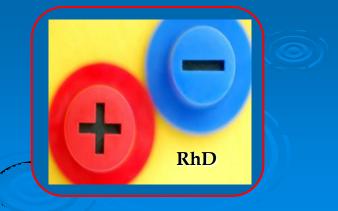
CELL-FREE FETAL DNA

Fetal gender determination Diagnostic accuracy: 99,8%



Fetal RhD genotyping Diagnostic accuracy: 97,5%



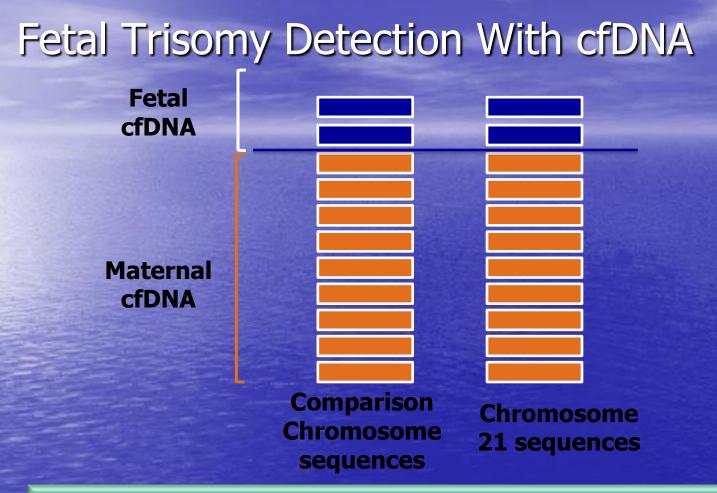


PRENATAL ASSESSMENT OF FETAL RhD STATUS

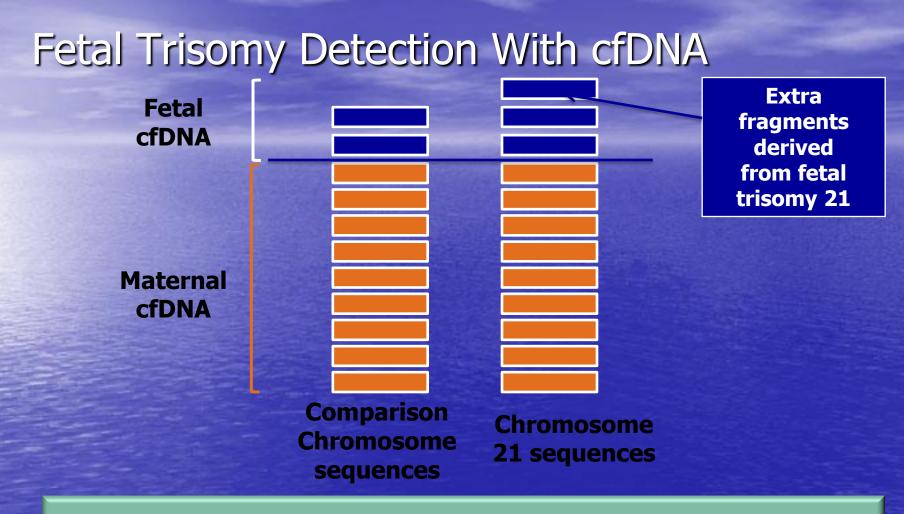
1. So far, the test has been performed on 166 pregnant women

2. The test is functional in clinical routine practice of non invasive prenatal diagnosis since it is easy, rapid and automated. After about 4 hours from the blood sampling it is possible to obtain the results of 20 samples simultaneously.

SENSITIVITY (%)	97.7
SPECIFICITY (%)	100
VPP (%)	100
VPN (%)	96.3
EFFICIENCY (%)	99.8



* Each bar represents hundreds of cfDNA fragments
 * Counting of chromosome cfDNA fragments done by DNA sequencing



* Overabundance of chromosome 21 cfDNA fragments in T21, although small, can be measured with DNA sequencing

Cell-free DNA in maternal blood Screening for aneuploidies

	FF 2	%	FF 4	%	FF 1	0%	FF	20%
Model Fetal DNA								
XIIXIX Maternal DNA				•••		•••		••• •••
	Euploid	T21	Euploid	T21	Euploid	T21	Euploid	

- Cell-free DNA (cfDNA) are short DNA fragments
- In pregnancy, cfDNA from both the mother and fetus are in maternal blood
- Amount of fetal cfDNA present is a small fraction of the maternal cfDNA

NIPT Technology Different approaches to cfDNA analysis

Massively Parallel Shotgun Sequencing (MPSS)

Directed Approach (e.g. Harmony Test)

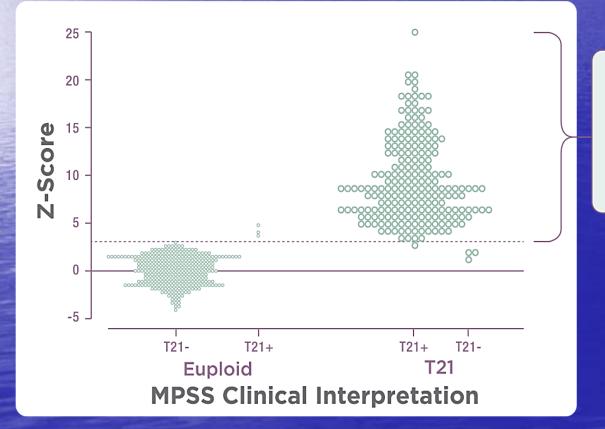
Key differences

Binary +/- result based on z-score

Risk classification and risk score

Massively Parallel Shotgun Sequencing (MPSS)

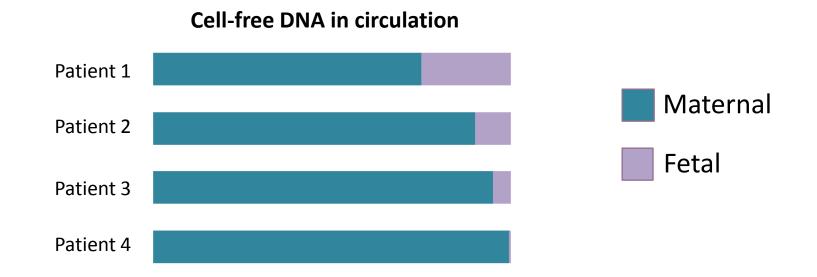
MPSS is a random sampling of cfDNA fragments
 An arbitrary z-score cut-off is used to determine trisomy



All classified as "positive" with no distinction between extremely high values and those just above the cut-off

Palomaki GE et al., Genet Med. 2011 Nov;13(11):913-20.

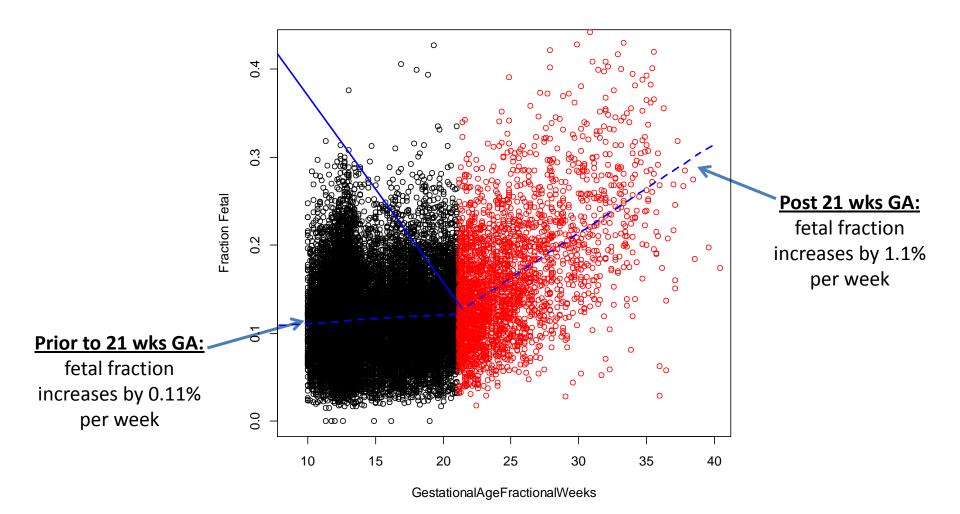
Importance of measuring fetal DNA amount



- Percentage of maternal to fetal DNA in circulation can vary from woman to woman, and changes throughout gestation¹
- In some samples, there is very little or no detectable fetal DNA
- Important to choose a lab that measures fetal fraction
 - CAP accreditation program recommends that NIPT labs measure and report fetal fraction²

1. Wang E et al, Prenat Diagn. 2013 Jul;33(7):662-6. 2. College of American Pathologists Molecular Pathology Checklist. MOL.34927. 7/29/2013

Fetal Fraction – Gestational Age Relationship



Consequences of NOT measuring fetal DNA

If very little fetal DNA is present, result is based on maternal DNA

- Male fetuses may be called as "female"
- PATIENTS CARRYING FETUS WITH TRISOMY MAY RECEIVE FALSE REASSURANCE (increased risk of "false negative" results)
- If fetal DNA percent is not measured and reported, validity of individual result is not known

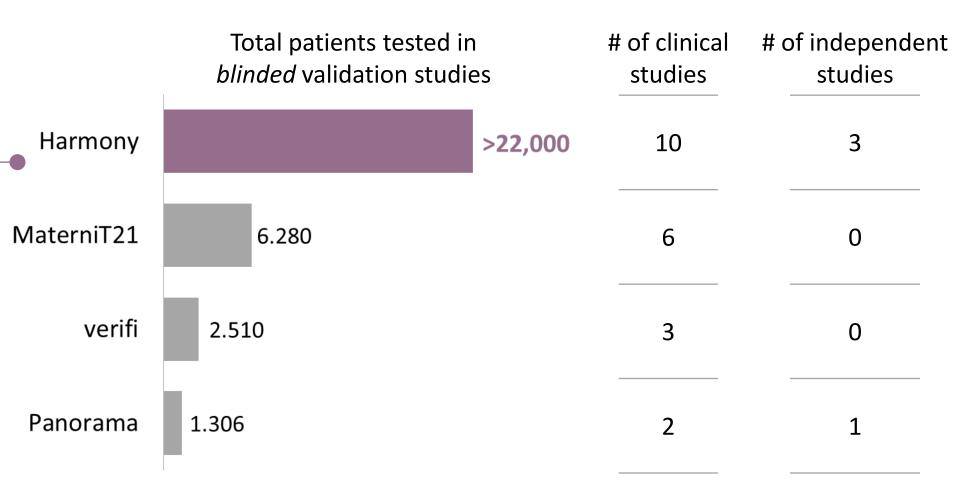
"Can deeper sequencing alleviate the need to measure fetal DNA?"

- A recent independent study¹ evaluated various depths of sequencing of chromosome 21 at varying levels of fetal fraction
 - Conclusion: Detection rates will suffer if fetal DNA amount is
 3% or less <u>at any depth of sequencing</u>

Comparison of NIPT Tests

	Harmony	MaterniT21+ (Sequenom)	verifi (Verinata)	NIFTY (BGI)	PraenaTest (Lifecodexx)	Panorama (Natera)
Technology	Directed	MPSS (random)	MPSS (random)	MPSS (random)	MPSS (random)	Directed
Fetal fraction measured for proper testing	+	+			+	+
Test success rate	+	+	+	+	+	_
Individualized risk score	+	_	_		_	+
Low cost	+	_	_	+	_	_
Robust clinical studies	+	+	_	_	_	_

Harmony is backed by extensive evidence



As of October 2014

Extensive Clinical Data

Clinical Validity and Use

Fetal Fraction

Study	Subjects	Reference
NEXT – General pregnancy, 1 st trimester	18,955	NEJM 2015
NICE - Cohort validation study	3,228	Norton M et al., AJOG 2012
Clinical experience in Belgium & Netherlands	3,000	Willems et al, FVV 2014
General pregnancy population, 1st trimester	2,049	Nicolaides et al, AJOG 2012
Trisomy 13	1,949	Ashoor et al., Ultra Obstet Gyn 2013
Kypros Nicolaides clinical implementation	1,005	Mar Gil et al, Ultra Obstet Gyn 2013
EU-NITE - European study	520	Verweij et al., Prenatl Diag, 2013
High-risk population, 1st trimester	400	Ashoor et al., AJOG 2012
FORTE	338	Sparks et al., AJOG 2012
DANSR	298	Sparks et al., Prenat Diagn 2012
Ob/Gyn real world experience	289	Fairbrother et al., Prenat Diagn 2013
Twins study	275	Mar Gil et al., Fetal Diagn Ther 2013
Sex chromosome aneuploidies, study 1	177	Nicolaides et al., Fetal Diagn Ther 2013
Sex chromosome aneuploidies, study 2	432	Hooks et al., Prenat Diagn 2014

Maternal weight effects - commercial data	22,000	Wang et al., Prenat Diagn 2013
Consistent in high and low-risk women	3,007	Brar et al, J Mat Fet Neonat Med 2013
Fetal cfDNA and pregnancy complications	1,949	Poon et al., Fetal Diagn Ther 2013
Maternal weight and fetal factors, study 2	1,949	Ashoor et al. Ultras Obstet Gyn 2013
Maternal weight and fetal factors, study 1	400	Ashoor et al., Fetal Diagn Ther 2012
Fetal fraction in twins	70	Struble et al., Fetal Diagn Ther 2013

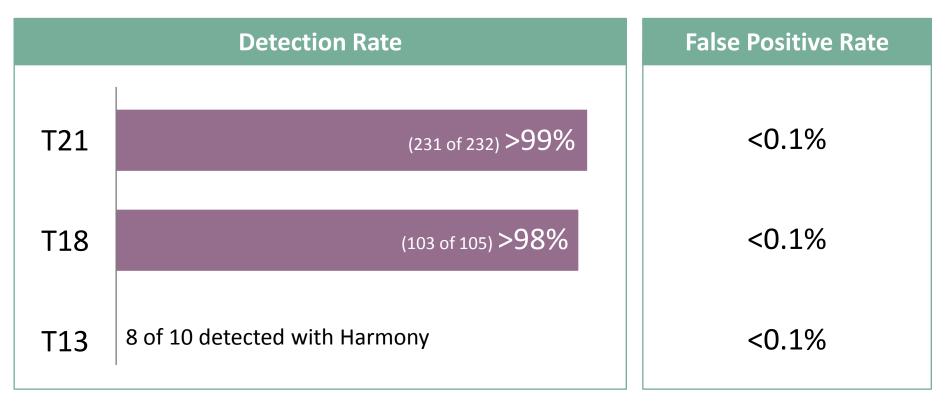
Cell free DNA test

Trisomy 21

Author		DR (95% CI)	Wt (%)			Wt (%)
Chiu et al., 2011 [86]		100.0 (95.8 to 100)	10.52		2.06 (0.43 to 5.9)	1.52
Ehrich et al., 2011 [39]		100.0 (91.0 to 100)	4.84		0.24 (0.01 to 1.4)	4.02
Palomaki et al., 2011 [212]		98.6 (95.9 to 99.7)	25.76		0.20 (0.04 to 0.6)	11.87
Sehnert et al., 2011 [13]		100.0 (75.0 to 100)	1.69		0.00 (0.00 to 10.3)	0.37
Ashoor et al., 2012 [50]	A REAL PROPERTY AND	100.0 (92.9 to 100)	6.17		0.00 (0.00 to 1.1)	3.45
Bianchi et al., 2012 [89]		100.0 (95.9 to 100)	10.88		0.00 (0.00 to 0.9)	3.97
Jiang et al., 2012 [16]		100.0 (79.4 to 100)	2.06		0.00 (0.00 to 0.4)	7.93
Lau et al., 2012 [11]		100.0 (71.5 to 100)	1.45		0.00 (0.00 to 3.7)	1.02
Nicolaides et al., 2012 [8]		100.0 (63.1 to 100)	1.09		0.00 (0.00 to 0.2)	14.54
Norton et al., 2012 [81]		100.0 (95.6 to 100)	9.92		0.04 (0.00 to 0.2)	18.89
Sparks et al., 2012 [36]		100.0 (90.3 to 100)	4.47		0.00 (0.00 to 2.8)	1.37
Zimmerman et al., 2012 [11]		100.0 (71.5 to 100)	1.45		0.00 (0.00 to 2.7)	1.40
Guex <i>et al.,</i> 2013 [30]		100.0 (88.4 to 100)	3.75		0.00 (0.00 to 2.5)	1.52
Liang et al., 2013 [37]		100.0 (90.5 to 100)	4.60		0.00 (0.00 to 1.0)	3.63
Nicolaides et al., 2013 [25]		100.0 (86.3 to 100)	3.14		0.00 (0.00 to 1.8)	2.09
Song et al., 2013 [8]		100.0 (63.1 to 100)	1.09		0.00 (0.00 to 0.2)	13.41
Stumm et al., 2013 [39]		97.4 (86.5 to 99.9)	4.84	-	0.00 (0.00 to 0.9)	4.20
Verweij <i>et al.,</i> 2013 [18]		94.4 (72.7 to 99.9)	2.30	The second se	0.00 (0.00 to 0.7)	4.83
Pooled analysis [809]	•			•		
	50 60 70 80 90 100 DR % (95% CI)			0 3 6 9 12 FPR % (95% Cl)	Gil et a	al., 2013

ANTA

High detection rate; low false positive rate



Studied in over 6,000 patients, including >2,000 average-risk women

Mosaicism

- 1. Sparks AB et al., Am J Obstet Gynecol. 2012 Apr;206(4):319.e1-9.
- 2. Ashoor G et al., Am J Obstet Gynecol. 2012 Apr;206(4):322.e1-5.
- 3. Sparks AB et al., Prenat Diagn. 2012 Jan;32(1):3-9.
- 4. Norton M et al., Am J Obstet Gynecol. 2012 Aug;207(2):137.e1-8.
- 5. Nicolaides KH et al., Am J Obstet Gynecol. 2012 Nov;207(5):374.e1-6.
- 6. Ashoor G et al., Ultrasound Obstet Gynecol. 2013 Jan;41(1):21-5.
- 7. Data on file

Screening for trisomy 21 1960-2013





International Federation of Gynecology and Obstetrics Working Group on Best Practice in Maternal-Fetal Medicine

Chair: G C Di Renzo

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SCREENING FOR CHROMOSOMAL ABNORMALITIES AND NON INVASIVE PRENATAL DIAGNOSIS AND TESTING

- Maternal age has a low performance as a screening for fetal chromosomal abnormalities with a DR of 30-50% for FPR of 5-20%. Therefore, invasive testing for diagnosis of fetal aneuploidies should not be carried out by taking into account only maternal age.
- First-line screening for trisomies 21, 18 and 13 should be achieved by the combined test, which takes into account maternal age, fetal nuchal translucency (NT) thickness, fetal heart rate (FHR) and maternal serum free β-human chorionic gonadotropin (β-hCG) and pregnancy-associated plasma protein-A (PAPP-A). The combined risk test has a DR of 90% for trisomy 21 and 95% for trisomies 18 and 13, at FPR of about 5%.

- The combined test could be improved by assessing additional ultrasonographic markers, including the fetal nasal bone and Doppler assessment of the fetal ductus venosus flow and tricuspid flow. If all those markers are included the DR is increased to more than 95% and the FPR decreased to less than 3%.
- Screening by analysis of cfDNA in maternal blood has a DR of 99% for trisomy 21, 97% for trisomy 18 and 92% of trisomy 13, at a total FPR of 0.4%.

- Clinical implementation of cfDNA testing should preferably be in a contingent strategy based on the results of first-line screening by the combined test at 11-13 weeks' gestation. In this case, we recommend the strategy below:
 - Combined test risk over **1** in **100**: the patients can be offered the options of cfDNA testing or invasive testing.
 - Combined test risk between **1** in **101** and **1** in **2,500**: the patients can be offered the option of cfDNA testing
 - Combined test risk lower than **1 in 2,500**: there is no need for further testing.



FETAL CELLS and ffDNA: CONCLUSIONS AND FUTURE PERSPECTIVES



Although new methodologies based on SNPs and free fetal nucleic acids are arising, we believe that the use of fetal cells is still a good approach for non invasive prenatal diagnosis of fetal trisomies, because it allows us to visualize directly the fetal nuclei and their chromosomes. *In this respect, our SAFE test is up to know the only one offered at clinical level.*

If DNA can be utilised with high specificity and sensitivity for the determination of fetal sex and fetal RhD status as early as 9 wks gestation. Moreover recently its applicability for the diagnosis of trisomies has been clearly validated.

